A HIGHLY ACTIVE CHEMOTACTIC PEPTIDE ANALOG INCORPORATING THE UNUSUAL RESIDUE 1-AMINOCYCLOHEXANECARBOXYLIC ACID AT POSITION 2

M. Sukumar 1, P. Antony Rajl, P. Balaram 1 and E. L. Becker 2

 Molecular Biophysics Unit, Indian Institute of Science, Bangalore 560 012, India
Department of Pathology, University of Connecticut Health Center, Farmington, CT 06032

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SUMMARY: Analogs of chemotactic peptides (Formyl-Met-X-Phe-OMe) containing the residues a-aminoisobutyric acid (Aib), constrained stereochemically aminocyclopentanecarboxylic acid (Acc5) and 1-aminocyclohexanecarboxylic acid (Acc6) at position 2 are compared with the parent sequence (X=Leu) for their ability to induce lysozyme release in rabbit neutrophils. The Acc6 analog is about 78 times more active than the parent peptide, For-Met-Leu-Phe-OH, whereas Aib and Acc⁵ analogs are approximately 3 and 2 times, respectively, less active than the parent peptide. NMR and model building studies clearly favour a Met-Acc6 8-turn solution conformation in the Acc⁶ analog, suggesting that the neutrophil receptor is capable of recognizing a folded peptide structure. The significant differences in the activities of the Acc⁵ and Acc⁶ analogs suggest an important role for the residue 2 sidechain in receptor interactions. © 1985 Academic Press, Inc.

The tripeptide Formyl-methionyl-leucyl-phenylalanine (For-Met-Leu-Phe-OH)⁺ induces neutrophil chemotaxis and release of lysosomal enzymes (l). These effects are undoubtedly mediated by specific interactions of the peptide with receptors on the neutrophil cell surface (l). Structure-activity correlations using a wide variety of chemotactic peptide analogs (l-3) have been interpreted in terms of extended β-sheet structures (4), as the 'biologically active' (receptor bound conformation) of the peptide (5). Recent studies using stereochemically constrained analogs, incorporating α-aminoisobutyric acid (Aib) or l-aminocyclopentane-carboxylic acid (Acc⁵, earlier referred to as cycloleucine, Cyl) however suggest that folded conformations are indeed biologically active (6). The folded analogs were found to be marginally less active than the parent peptide. This could be due to less effective receptor recognition of the folded backbone conformation or a consequence of less than optimal interactions of the residue 2 sidechain

Abbreviations:

Acc^{η}, l-aminocycloalkanecarboxylic acid residue, where n is the number of atoms in the cycloalkane ring; Aib, α -amino-isobutyric acid; Boc, t-butyloxycarbonyl; For, formyl; TEMPO, 2, 2, 6, 6-tetramethyl-piperidine-l-oxyl.

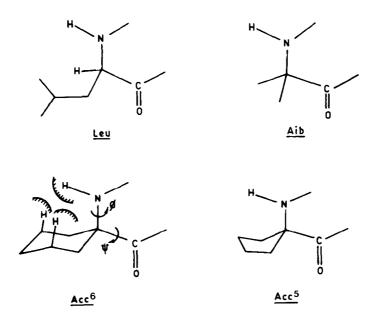


Figure 1 Structures of sidechains at position 2. Note 1, 3-diaxial interactions in Acc⁶ restrict 0 values.

at the receptor site. In order to distinguish between these possibilities we examined the peptide For-Met-Acc⁶-Phe-OMe, in which the residue 2 sidechain more closely approximates the Leu sidechain (Figure 1). This report establishes that the Acc^6 analog is significantly more active than the parent peptide and presents evidence for a folded β -turn solution conformation.

MATERIALS AND METHODS

1-aminocyclohexanecarboxylic acid (H-Acc⁶-OH) was synthesized from cyclohexanone, using a procedure analogous to that described for α-aminoisobutyric acid (7). All peptides were synthesized by conventional racemization free procedures, using Boc and methyl ester groups for amino and carboxyl protection. Peptides were purified by column chromatography on silica gel and shown to be homogeneous by reverse phase HPLC on a Lichrosorb RP-18 column using methanol-water gradient elution (40-70% methanol in 30 min, flow rate 0.8 ml min⁻¹). The sample of For-Met-Acc⁶-Phe-OMe was also purified by HPLC. The peptides were fully characterized by 270 MHz ¹H NMR. All NMR studies were carried out on a Varian FT-80A spectrometer or a Bruker WH-270 spectrometer at the Sophisticated Instruments Facility, Bangalore. Lysozyme release from cytochalasin B treated rabbit neutrophils was measured as described earlier (1).

RESULTS AND DISCUSSION

Table 1 summarizes the granule enzyme releasing activity of four chemotactic peptide analogs, For-Met-X-Phe-OMe (X = Leu $\underline{1}$, Aib $\underline{2}$, Acc⁵ $\underline{3}$ and Acc⁶ $\underline{4}$). The peptides $\underline{2}$ and $\underline{3}$ possess significant activity, whereas the Acc⁶ analog 4 is approximately 12.5 fold more

Peptide	ЕD ₅₀ (м) ^а	Relative Activity b	Met NH		NH NMR Par X -	NH NMR Parameters ^b X - NH	Phe NH	HN
			ds/dT ∆s		d8/dT ∆6	Δδ	ds/dT	Δδ
X = Leu]	$7.4 \pm 0.5 \times 10^{-11}$ (7)	6.2	6.2	1.38	5.1	0.92	4.3	1.36
X = Aib 2	$7.4 \pm 0.6 \times 10^{-10}$ (4)	0.32	5.0	1.65	4.7	~.	3.5	0.93
$X = Acc^5 \frac{3}{3}$	$4.7 \pm 0.6 \times 10^{-10}$ (4)	0.51	5.2	2.15	4.7	1.55	3.2	0.65
$X = Acc^6 4$	$5.0 \pm 0.6 \times 10^{-12}$ (4)	7.8	5.3	2.2	4.8	1.47.	3.3	0.73

The figure in the parentheses is the number of independent observa-ED50 is the peptide concentration required to produce 50% of the maximum effect determined by the concentration-Each value is the average ± SE. effect curve. tions.

periment, the ED50 of the reference peptide For-Met-Leu-Phe-OH was determined. The mean ED50 ± S.E. of For-Met-Leu-Relative to For-Met-Leu-Phe-OH. The activities of all of the peptides were not measured at the same time and in the To take account of possible differences in the sensitivity of the cells from experiment to ex-Phe-OH determined in the same experiment as the given peptide methyl ester (figure in parenthesis following the ED50) are: $4.6 \pm 0.5 \times 10^{-10} M$ (1); $2.4 \pm 0.5 \times 10^{-10}$ (2, 3); $3.9 \pm 0.7 \times 10^{-10} M$ (4). same experiments. Q

 $d\delta/dT$ is the temperature coefficient (ppm/K \times 10³) measured in (CD₃)₂ SO. $\Delta\delta$ = δ [(CD₃)₂SO₃- δ (CDCl₃) ppm, (c)

active than the Leu analog 1. The peptide esters 1, 2, 3 have higher activity than the corresponding peptide acids (6, see also ref. 3). For-Met-Acc⁶-Phe-OMe is the first tripeptide analog exhibiting substantially greater activity than the parent peptide, although enhanced activity has been very recently reported for tetrameric analogs having four peptide molecules covalently linked to a macrocyclic tetraazacycloalkane (8).

The high biological activity of For-Met-Acc6-Phe-OMe provides a strong stimulus for determining its solution conformation. Solvent shielded or intramolecularly hydrogen bonded NH groups in the peptide were delineated by a comparison of the temperature dependence of chemical shifts in (CD3)SO, solvent dependence of NH chemical shifts in CDCl3-(CD3)2SO mixtures and paramagnetic radical induced line broadening in CDCl3 (9). The temperature coefficient (d δ/dT) and solvent shift ($\Delta \delta$) values for the four peptides are compared in Table 1. It is clearly seen that the Met and Leu NH groups are significantly more sensitive to changes in solvent composition or temperature, as compared to the Phe NH group in peptides $\underline{2} - \underline{4}$. In $\underline{1}$, all three NH groups show comparable sensitivity. A similar observation is made in the free radical induced line broadening experiment. Addition of 2, 2, 6, 6-tetramethylpiperidine-l-oxyl (TEMPO) to a CDCl3 solution of 1 resulted in almost equal broadening of all three NH groups. On the contrary, in peptides 2 - 4 the order of line broadening is Met NH > X NH >> Phe NH. The NMR results thus strongly suggest that the Phe NH is appreciably shielded from the solvent in peptides 2.4. A detailed comparison of peptides 1 and 2 over a wide range of concentrations has led to a similar conclusion (10). It is therefore likely that the Phe NH is involved in an intramolecular hydrogen bond, since steric shielding of the NH group is unlikely in a small peptide. Supporting evidence for the presence of intramolecularly hydrogen bonded conformations comes from IR studies in CHC13, where an NH stretching band is seen at 3380-3390 cm⁻¹ even in very dilute solutions.

An examination of molecular models and preliminary conformational energy calculations suggests that the Acc^6 residue is stereochemically restricted almost exclusively to the right or left handed helical ($\phi = \mp 60 \pm 20^{\circ} \psi = \mp 30^{\circ} \pm 20^{\circ}$) regions of the conformational map. This conformational preference is also shown by the Aib and Acc^5 residues (II). However, in the Acc^6 residue, 1-3 diaxial interactions between the axial substituent at C-1 and the axial hydrogens at C-3 and C-5 further restricts the

Figure 2. β -turn conformation proposed for For-Met-Acc⁶ - Phe-OMe.

accessible values of ϕ . It should be noted that the amino group in Acc^6 necessarily adopts the axial position (12), a feature observed in two recent crystal structures of model Acc^6 peptides (13). In the crystal conformations of Boc-Aib-Acc⁶-OMe and Boc-Aib-Acc⁶-NHMe the conformational angles observed for the Acc^6 residue are $\phi = \pm 48^\circ \psi = \pm 42.6^\circ$ and $\phi = \pm 68.4^\circ \psi = \pm 15^\circ$, respectively (13).

These observations suggest that either a Type I or Type II-Met-Acc⁶- β -turn conformation (Figure 2) is consistent with the NMR data for peptide $\underline{4}$ and the known stereochemical preferences for the Acc^6 residue. Although a distinction between these structures is not possible on the basis of the available data, the results provide compelling evidence for the high biological activity of folded, conformationally constrained chemotactic peptide analogs. The dramatic enhancement in activity observed by introduction of a single methylene group in the position 2 sidechain in the Acc^6 peptide as compared to the Acc^5 analog, strongly suggests that peptide-receptor interactions involving this site are an important determinant of biological effects.

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REFERENCES

- Showell, H.J., Freer, R.J., Zigmond, S.H., Schiffmann, E., Aswanikumar, S., Corocran, B.A. and Becker, E.L. (1976) J. Exp. Med. 143, 1154-1169.
- Freer, R.J., Day, A.R., Radding, J.A., Schiffmann, E., Aswanikumar, S., Showell, H.J. and Becker, E.L. (1980) Biochemistry 19, 2040-2410.
- 3. Toniolo, C., Bonora, G.M., Showell, H.J., Freer, R.J. and Becker, E.L. (1984) Biochemistry 23, 698-704.
- 4. Becker, E.L., Bleich, H.E., Day, A.R., Freer, R.J., Glasel, J.A. and Visintainer, J. (1979) Biochemistry 18, 4656-4668.
- 5. Freer, R.J., Day, A.R., Muthukumaraswamy, N., Pinon, D., Wu, A., Showell, H.J. and Becker, E.L. (1982) Biochemistry 21, 257-263.
- Iqbal, M., Balaram, P., Showell, H.J., Freer, R.J. and Becker, E.L. (1984) FEBS Letters 165, 171-174.
- 7. Clarke, H.T. and Bean, H.J. (1943) Organic Synthesis Coll. Vol. 2, 29-31.
- 8. Kraus, J.L., Dipaola, A. and Bellean, B. (1984) Biochem. Biophys. Res. Commun. 124, 939-944.
- Wuthrich, K. (1976) NMR in Biological Research: Peptides and Proteins, Elsevier, Amsterdam.
- 10. Raj, P.A. and Balaram, P. (1985) Biopolymers. In press.
- 11. Prasad, B.V.V. and Balaram, P. (1984) CRC Crit. Revs. Biochem. 16, 307-348.
- 12. Kenner, G.W., Preston, J. and Sheppard, R.C. (1965) J. Chem. Soc. 6239-6244.
- Bardi, R., Piazzesi, A.M., Toniolo, C., Sukumar, M., Raj, P.A. and Balaram, P. (1985) Int. J. Peptide Protein Res. In press.